

AMENDMENTS TO THE SPECIFICATION

Please replace the last paragraph on page 54 as follows:

GPA_{ID} and GPA_{LW} hybrids. The regions of high homology among G α subunits that have been identified by sequence alignment are interspersed throughout the molecule. The G1 region containing the highly conserved -GSGESGDST- (SEQ ID NO:127) motif is followed immediately by a region of very low sequence ~~conservation~~ conservation, the "i1" or insert 1 region. Both sequence and length vary considerably among the i1 regions of the G α subunits. By aligning the sequences of G α subunits, the conserved regions bounding the i1 region were identified and two additional classes of GPA1-G α hybrids were constructed. The GPA_{ID} hybrids encode the amino terminal 102 residues of GPA1 (up to the sequence -QARKLGIQ- (SEQ ID NO:122)) fused in frame to mammalian G α subunits, while the GPA_{LW} hybrids encode the amino terminal 244 residues of GPA1 (up to the sequence -LIHEDIKA- (SEQ ID NO:123) in GPA1). The reason for constructing the GPA_{ID} and GPA_{LW} hybrids was to test the hypothesis that the i1 region of GPA1 is required for mediating the interaction of GPA1 with yeast G $\beta\gamma$ subunits, for the stable expression of the hybrid molecules, or for function of the hybrid molecules. The GPA_{ID} hybrids contain the amino terminal domain of GPA1 fused to the i1 domain of mammalian subunits, and therefore do not contain the GPA1 i1 region, while the GPA_{LW} hybrids contain the amino terminal 244 residues of GPA1 including the entire i1 region (as defined by sequence alignments). Hybrids of both GPA_{ID} and GPA_{LW} classes were constructed for G α S, G α i2, G α i3, G α o_a, and G α 16; none of these hybrids complemented the *gpa1* growth arrest phenotype.

Please replace the last paragraph on page 55 and the first paragraph on page 56 as follows:

G α s Hybrids. There is evidence that the "switch region" encoded by residues 171-237 of G α transducin (using the numbering of Noel et al (1993)) also plays a role in G $\beta\gamma$ coupling. First, the G226A mutation in G α S (Miller et al. 1988) prevents the GTP-induced conformational change that occurs with exchange of GDP for GTP upon receptor activation by ligand. This residue maps to the highly conserved sequence -DVGGQ- (SEQ ID NO:128), present in all G α subunits and is involved in GTP hydrolysis. In both the G α t and G α i1 crystal structures, this sequence motif resides in the loop that connects the β 3 sheet and the α 2 helix in the guanine nucleotide binding core. In addition to blocking the conformational change that occurs upon GTP binding, this mutation also prevents dissociation of GTP-liganded G α s from G $\beta\gamma$. Second, crosslinking data reveals that a highly conserved cysteine residue in the α 2 helix (C215 in G α o, C210 in G α t) can be crosslinked to the carboxy terminal region of G β subunits. Finally, genetic evidence (Whiteway et al. 1993) identifies an important single residue in GPA1 (E307) in the β 2 sheet of the core structure that may be in direct contact with $\beta\gamma$. A mutation in the GPA1 protein at this position suppresses the constitutive signalling phenotype of a variety of STE4 (G β) dominant negative mutations that are also known to be defective in G α -G $\beta\gamma$ association (as assessed in two-hybrid assay in yeast as well as by more conventional genetic tests).

Please replace the paragraph in the middle of page 98 as follows:

Clearly, these sequences encode novel peptides, as the native α -factor sequence differs considerably:

Tyr Ile Ile Lys Gly Val Phe Trp Asp Pro Ala (SEQ ID NO:129).

Please replace the full paragraph on page 110 as follows:

A hybrid gene encoding the prepro-region of human POMC (accession # K02406; Takahashi, H., et al (1983) Nucleic Acids Research 11:6847-6858) and the coding region of a single repeat of mature α -factor will be constructed in the following fashion. The prepro-region of human POMC will be amplified with an HindIII site at the 5' end and a BbsI site at the 3' end using VENT polymerase and the following primers: 5' GGGAAGCTT **ATGCCGAGATCGTGCTGCCAGCCGC** 3' (SEQ ID NO:30) (HindIII site is underlined and initiation codon is italic bold) and antisense 5' GGGGAAGACTTCTGCCCTGCGCCGCTGCTGCC 3' (SEQ ID NO:31) (BbsI recognition is underlined), leaving the amino acid sequence -SSGAGQ**KR**- (SEQ ID NO:125) at the 3' end with a Bbs1 site leaving an overhang at the -KR- dibasic cleavage sequence. The coding region of α -factor will be amplified from Cadus 1219 with a Bbs1 site at the 5' end and a BglII site at the 3' end using the primers 5' GGGGAAGACCCGCAGGAGGCAGAAGCTT GGTTCAG 3' (SEQ ID NO:32) (BbsI site is underlined) and 5' GGGAGATCTTCAGTACATTGGTTGGCC 3' (SEQ ID NO:33) (BglII site is underlined, termination codon is bold). The PCR fragment encoding the pre-pro segment of POMC is restricted with HindIII and BbsI and gel purified, the PCR fragment encoding α -factor is cut with BbsI and BglII and gel purified, and Cadus 1215 is cut with BglII and partially with HindIII and the HindIII-BglII restricted vector containing the pAlter polylinker sequences is gel purified. Three-part ligation of the two PCR products with HindIII and BglII digested Cadus 1215 will yield a hybrid POMC/ α -factor gene in which the first 104 amino acids residues are from POMC and the remaining 17 are from α -factor. The structure of this hybrid gene around the PC1 cleavage site is: -RNSSSSGSSGAGQ**KREAEAWHWLQLKPGQPMY*** (SEQ ID NO:34) where residues donated by POMC are underlined, the dibasic cleavage site is underlined bold, and the sequence of mature α -factor is in italics. The tetrapeptide -EAEA- (SEQ ID NO:130) juxtaposed between the dibasic cleavage site and the amino-terminal tryptophan of mature α -factor should be removed by the dipeptidyl aminopeptidase activity of ste13p.

Please replace the first line of Table 7 on page 153 as follows:

GPA1 RIDTTGITETEFNIGSSKFKVLDAGGQSRERKKWIHCFEGITAV
LFVLAMSEYDQMLFEDER (SEQ ID NO:131)